

An Effect of Light Intensity on the Total Flavonoid and Phenolic Content of *Moringa Oleifera*

Aisha Idris^{1,2}, Alona C. Linatoc¹, Surayya M. Muhammad² and Zakiyyu Ibrahim Takai³

¹Faculty of Applied Sciences and Technology, Universiti Tun Hussein Onn Malaysia (UTHM), Hub Pendidikan Tinggi Pagoh, KM1, Jalan Panchor, 84600, Muar, Johor, Malaysia

²Faculty of Sciences, Federal University Dutse, PMB 7156, Jigawa State, Nigeria

³Yusuf Maitama Sule University, PMB 3236, Kano State, Nigeria

Abstract

Flavonoid and phenolic compounds are secondary metabolites produced by plants in response to harsh environmental conditions. Light is one of the most important ingredients that affects their production. *Moringa oleifera* (also known as *M. oleifera*), a plant belonging to the family *Moringaceae* is having bioactivity due to the flavonoid and phenolic contents of the plant. The objective of the research is to determine the effect of varying light intensity on the total flavonoid and total phenolic contents of *M. oleifera*. Total flavonoid contents was determine based on the aluminium chloride colourimetry method while total phenolic contents were determined based on the folin-chiocalteau reagent. The results obtained in the study shows that the flavonoid and phenolic contents recovered from sun exposed plants was higher than that of shaded plants. Besides, comparison of the means showed that there is a statistically significant difference between the flavonoid and phenolic contents of the studied plant ($P < 0.05$). This leads to a conclusion that high light intensity can increase the concentration of flavonoid and phenolic contents of *M. oleifera*.

Keywords: Flavonoid, light intensity, *Moringa oleifera*, and phenolic content.

1. Introduction

Moringa oleifera is a drought-resistant tree belonging to the family *Moringaceae*. This fast-growing deciduous tree is widely cultivated in a tropical region. It produces edible pods and leaves which are having high application in traditional medicine. Moreover, the plant can be used in water purification. Numerous researchers reported the medicinal benefit of the tree. The antioxidant activity [1,2,3,4,5,6,7, and 8], photoprotective effect [9], bioactive flavonoids [10], immunomodulatory effect [11], antidiabetic [12], antibacterial [13], anticancer [14], antiproliferative effect [15] and antiulcerogenic effect [16] were reported.

Phenolic compounds and flavonoids are plant secondary metabolites produced as a response to harsh environmental condition. They function as UV filters, antimicrobial agents, and

pollinator attractants. The availability of phenolic and flavonoid in a plant makes a plant useful for healing and curing diseases [29]. Light is an important environmental factor that can affect the accumulation of phenolic and flavonoid contents of a plant. This is possible because every plant has a specific light requirement for its maximum production of flavonoid and phenolic compounds. For instance, *Lithocarpus litseifolius* [17] and *Zingiber officinale* [18] accumulates high amount of flavonoid under lower light intensity while Glycine max seedling [19] produces a high amount of flavonoids under high light intensity. *Labisa pumila* accumulates more phenolic and flavonoid under lower light intensity [20]. *Anoectochilus roxburghii* produced a high amount of flavonoid under blue light and high amount of phenolic under red film [21]. Due to these variations, there is a need to know the level of light intensity that leads to a maximum accumulation of flavonoid and phenolic contents of *M. oleifera*. As such, the objective of this research is to evaluate the effect of varying light intensity on the total phenolic and flavonoid contents of *M. oleifera*.

2. Methodology

2.1 Plant Material

Sun exposed and shaded *M. oleifera* plants were sampled randomly. Leaves were harvested and dried. Extraction was done according to Chai et al. [22] using ethanol as a solvent. The plant extract was then stored at lower temperature and later used for the determination of total flavonoid and phenolic contents of the plant.

2.2 Determination of the Total Flavonoid Content

This was achieved using the colourimetry method. The plant extract was mixed with 10% aluminium chloride and 1M potassium acetate. Absorbance was recorded at 420nm. Quercetin was used as a standard and the result was expressed as mg/g quercetin equivalent [23].

2.3 Determination of the Total Phenolic Content

The total phenolic content was determined spectrophotometrically using Folin-chiocalteau reagent. The plant extract was mixed with the reagent and NaCO₃. Absorbance was recorded at 730nm. Gallic acid was used as a standard and the result was expressed as mg/g Gallic acid equivalent [24].

2.4 Statistical analysis

SPSS statistical software was used to analyse the data. Differences in the means of the total phenolic and flavonoid contents of sun exposed and shaded plants were compared using T-Test at 95% confidence level. Results were given as mean plus or minus standard deviation.

3. Result and Discussion

The total flavonoid and phenolic content of *M. oleifera* are represented in Figure 1. From the result obtained, it is seen that leaves of sun exposed plant were having higher flavonoid and phenolic contents than leaves of the shaded plant. This may be due to the photoprotective role of flavonoid and phenolics. Comparison of means showed that there is a statistically significant difference between the total flavonoid and phenolic content of sun exposed and shaded *M. oleifera* ($P < 0.05$).

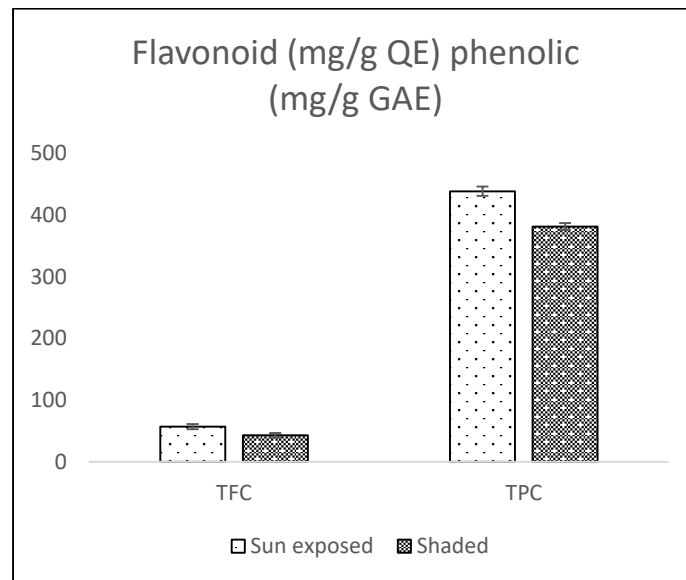


Figure 1. The total flavonoid and phenolic content of sun exposed and shaded *M. oleifera*

The total flavonoid content of sun exposed plant was found to be 57mg/g QE FW. In another study, the total flavonoid content of *M. oleifera* was found to be 90mg/g QE [25], about 80mg/g QE of flavonoid was recovered by [14], 13g/100g QE [3], and 27mg/g QE [8]. The total phenolic content of sun exposed plant was found to be 438 mg/g GAE FW. In another study, the phenolic content recovered from *M. oleifera* was found to be about 2000mg/g GAE [25], 900 mg/g [14], 1460mg/g [13] and 52 $\mu\text{g}/\text{mg}$ [9].

Many plants produce a high amount of phenolic content under high light intensity. For instance *Zingiber officinale* [18], *Zea mays* sprouts [26], *Labisa pumila* [20], *Glycine max* sprout [19] and *Gracila pumila* [24] requires a high amount of light for a maximum accumulation of

phenolics. On the other hand, flavonoid accumulation can be high in shaded or sun exposed plant. Some plants, for example, *Piper aduncum* [27] and *Zingiber officinale* [18] requires a moderate amount of shading for their maximum accumulation of flavonoid. besides, *Centella asiatica* [28] requires a sunny condition for their maximum accumulation of flavonoid.

4. Conclusion

The light intensity can affect the accumulation of flavonoid and phenolic contents of *M. oleifera*. The phenolic content recovered from the plant exceeds the flavonoid content recovered. Moreover, sun exposed plant was having higher amount of flavonoid and phenolic contents than shaded *M. oleifera*.

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References

- [1] S. Lalas and J. Tsaknis, "Extraction and Identification of Natural Antioxidant from the Seeds of the *Moringa oleifera* Tree Variety of Malawi," *JAOCS*, vol. 79, no. 7, pp. 677–683, 2002.
- [2] I. Karageorgou, S. Grigorakis, S. Lalas, and D. P. Makris, "Enhanced extraction of antioxidant polyphenols from *Moringa oleifera* Lam . leaves using a biomolecule - based low - transition temperature mixture," *Eur. Food Res. Technol.*, pp. 1–10, 2017.
- [3] S. I. Ã and M. I. Bhangar, "Effect of season and production location on antioxidant activity of *Moringa oleifera* leaves grown in Pakistan," *J. Food Compos. Anal.*, vol. 19, pp. 544–551, 2006.
- [4] M. Sarkar, S. Bhowmick, J. Hussain, M. Hasan, and S. Hossain, "Hot Water Extract of *Moringa oleifera* Leaves Protects Erythrocytes from Hemolysis and Major Organs from Oxidative Stress in vitro," *J. basic Appl. Res.*, vol. 3, no. 3, pp. 120–126, 2017.
- [5] G. Batra, O. Gortzi, S. I. Lalas, A. Galidi, A. Alibade, and G. D. Nanos, "Enhanced Antioxidant Activity of *Capsicum annum* L . and *Moringa oleifera* L . Extracts after Encapsulation in Microemulsions," *Chemengineering*, vol. 1, no. 15, pp. 1–13, 2017.
- [6] A. B. Falowo *et al.*, "Antioxidant activities of *Moringa oleifera* L . and *Bidens pilosa* L . leaf extracts and their effects on oxidative stability of ground raw beef during refrigeration storage," *CyTA - J. Food*, pp. 1–8, 2016.
- [7] M. M. Hasan, H. F. Alharby, A. S. Hajar, K. R. Hakeem, Y. Alzahrani, and S. Arabia, "Effects of magnetized water on phenolic compounds , lipid peroxidation and antioxidant activity of *moringa* species under drought stress," *J. Anim. plant Sci.*, vol. 28, no. 3, pp. 1–6, 2018.
- [8] S. Sreelatha and P. R. Padma, "Antioxidant Activity and Total Phenolic Content of *Moringa oleifera* Leaves in Two Stages of Maturity," *Plant foods Hum. Nutr.*, vol. 64, pp. 303–311, 2009.
- [9] A. B. Id *et al.*, "*Moringa oleifera* Leaf Extracts as Multifunctional Ingredients for ' Natural and Organic ' Sunscreens and Photoprotective Preparations," *Molecules*, vol. 23, no. 664, pp. 1–16, 2018.
- [10] M. Lin, J. Zhang, and X. Chen, "Bioactive fl avonoids in *Moringa oleifera* and their health-promoting properties," *J. Funct. Foods*, vol. 47, no. April, pp. 469–479, 2018.

- [11] V. Sai, A. Raju, N. Suryadevara, L. I. M. L. I. Chee, and N. E. Ismail, "Phytochemical analysis and immuno-modulatory effect of *Moringa oleifera*," *Int. J. Pharm. Pharm. Sci.*, vol. 9, no. 6, pp. 24–28, 2017.
- [12] G. A. Bamagous, S. S. Al Ghamdi, I. Abdel, A. Ibrahim, A. M. Mahfoz, and M. A. Afify, "Asian Pacific Journal of Tropical Biomedicine Antidiabetic and antioxidant activity of ethyl acetate extract fraction of *Moringa oleifera* leaves in streptozotocin-induced diabetes rats via inhibition of inflammatory mediators," *Asian Pac. J. Trop. Biomed.*, vol. 8, no. 6, pp. 320–327, 2018.
- [13] C. J. Guillén-román, R. G. Guevara-gonzález, N. E. Rocha-guzmán, A. Mercado-luna, and M. C. I. Pérez-pérez, "Industrial Crops & Products Effect of nitrogen privation on the phenolics contents , antioxidant and antibacterial activities in *Moringa oleifera* leaves," *Ind. Crop. Prod.*, vol. 114, no. March 2017, pp. 45–51, 2018.
- [14] L. Cuellar-Nuñez, I. Luzardo-Ocampo, R. Campos-Vega, M. A. Gallegos-Corona, E. González de Mejía, and G. Loarca-Piña, "Physicochemical and nutraceutical properties of moringa (*Moringa oleifera*) leaves and their effects in an in vivo AOM/ DSS-induced colorectal carcinogenesis model," *Food Res. Int.*, vol. FRIN 7126, pp. 1–45, 2017.
- [15] J. Tragulpakseerojn *et al.*, "Anti-proliferative effect of *Moringa oleifera* Lam (Moringaceae) leaf extract on human colon cancer HCT116 cell line," *Trop. J. Pharm. Res.*, vol. 16, no. 2, pp. 371–378, 2017.
- [16] D. Dahiru, J. Onubiyi, and H. A. Umaru, "Phytochemical screening and antiulcerogenic effect of *moringa oleifera* aqueous leaf extract," *Afr. J. Tradit. Complement. Altern. Med.*, vol. 3, no. 3, pp. 70–75, 2006.
- [17] A. Li *et al.*, "Effect of Light Intensity on Leaf Photosynthetic Characteristics and Accumulation of Flavonoids in *Lithocarpus litseifolius* (Hance) Chun. (Fagaceae)," *Open J. For.*, vol. 6, no. 5, pp. 445–459, 2016.
- [18] A. Ghasemzadeh, H. Z. E. Jaafar, and A. Rahmat, "Synthesis of phenolics and flavonoids in ginger (*Zingiber officinale* Roscoe) and their effects on photosynthesis rate," *Int. J. Mol. Sci.*, vol. 11, no. 11, pp. 4539–4555, 2010.
- [19] M. Yuan *et al.*, "Effect of fluorescence light on phenolic compounds and antioxidant activities of soybeans (*Glycine max* L. Merrill) during germination," *Food Sci. Biotechnol.*, vol. 24, no. 5, pp. 1859–1865, 2015.
- [20] E. Karimi, H. Z. E. Jaafar, A. Ghasemzadeh, and M. H. Ibrahim, "Light intensity effects on production and antioxidant activity of flavonoids and phenolic compounds in leaves, stems and roots of three varieties of *Labisia pumila* benth," *Aust. J. Crop Sci.*, vol. 7, no. 7, pp. 1016–1023, 2013.
- [21] S. Ye *et al.*, "Effects of Light Quality on Morphology, Enzyme Activities, and Bioactive Compound Contents in *Anoectochilus roxburghii*," *Front. Plant Sci.*, vol. 8, no. 5, pp. 1–7, 2017.
- [22] T. Chai, S. Elamparuthi, A. Yong, Y. Quah, and H. Ong, "activities of selected highland ferns of Malaysia," *Bot. Stud.*, vol. 54, no. 55, pp. 1–7, 2013.
- [23] V. Y. A. Barku, Y. Opoku-Boahen, E. Owusu-Ansah, and E. F. Mensah, "Antioxidant activity and the estimation of total phenolic and flavonoid contents of the root extract of *Amaranthus spinosus*," *Asian J. Plant Sci. Res.*, vol. 3, no. 1, pp. 69–74, 2013.
- [24] E. Cruces, M. R. Flores-Molina, M. J. Díaz, P. Huovinen, and I. Gómez, "Phenolics as photoprotective mechanism against combined action of UV radiation and temperature in the red alga *Gracilaria chilensis*," *J. Appl. Phycol.*, vol. 30, no. 2, pp. 1247–1257, 2017.
- [25] F. Ghafar *et al.*, "Total Phenolic Content And Total Flavonoid Content In *Moringa Oleifera*" *Sci. Herit. J.*, vol. 1, no. 1, pp. 23–25, 2017.
- [26] N. Xiang, X. Guo, F. Liu, Q. Li, J. Hu, and C. S. Brennan, "Effect of light- and dark-germination on the phenolic biosynthesis, phytochemical profiles, and antioxidant activities in sweet corn (*Zea mays* L.) sprouts," *Int. J. Mol. Sci.*, vol. 18, no. 6, pp. 1–13, 2017.
- [27] F. V. Pacheco, H. R. de Oliveira Silveira, A. A. Alvarenga, I. C. A. Alvarenga, J. E. B. P. Pinto, and J. M.

- S. Lira, "Gas Exchange and Production of Photosynthetic Pigments of *Piper aduncum* L. Grown at Different Irradiances," *Am. J. Plant Sci.*, vol. 4, no. December, pp. 114–121, 2013.
- [28] V. Müller, A. Albert, J. Barbro Winkler, C. Lankes, G. Noga, and M. Hunsche, "Ecologically relevant UV-B dose combined with high PAR intensity distinctly affect plant growth and accumulation of secondary metabolites in leaves of *Centella asiatica* L. Urban," *J. Photochem. Photobiol. B Biol.*, vol. 127, pp. 161–169, 2013.
- [29] A. Yahaya, M. Ali, and A. Idris. "Antibacterial activity and phytochemical screening of *Carica papaya* on some enteric bacterial isolates of public health importance." *Greener J of Biol Sci*, vol. 7, no. 1 pp. 001-007, 2017.